



# X-ray induced photodynamic therapy with copper-cysteamine nanoparticles in mice tumors

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**Photodynamic therapy (PDT), a treatment that uses a photosensitizer, molecular oxygen, and light to kill target cells, is a promising cancer treatment method. However, a limitation of PDT is its dependence on light that is not highly penetrating, precluding the treatment of tumors located deep in the body. Copper-cysteamine nanoparticles are a new type of photosensitizer that can generate cytotoxic singlet oxygen molecules upon activation by X-rays. In this paper, we report on the use of copper-cysteamine nanoparticles, designed to be targeted to tumors, for X-ray-induced PDT. In an in vivo study, results show a statistically significant reduction in tumor size under X-ray activation of pH-low insertion peptide-conjugated, copper-cysteamine nanoparticles in mouse tumors. This work confirms the effectiveness of copper-cysteamine nanoparticles as a photosensitizer when activated by radiation and suggests that these Cu-Cy nanoparticles may be good candidates for PDT in deeply seated tumors when combined with X-rays and conjugated to a tumor-targeting molecule.**

copper-cysteamine | radiotherapy | photosensitization | nanoparticles | cancer

Photodynamic therapy (PDT) involves the use of light and a photosensitizer that induces the production of reactive oxygen species at the tumor site after the absorption of light energy to kill nearby tumor cells (1–13). Singlet oxygen has a short lifetime in biological systems, less than 0.04  $\mu$ s, and therefore has a short radius of action of less than 0.02  $\mu$ m (10, 14). Thus, PDT is minimally invasive, and when used with light and a photosensitizer to selectively target cancerous cells, can minimize side effects to surrounding healthy tissues (1, 2, 5, 7, 10, 15–17). PDT is also unlikely to cause genotoxicity, rarely leads to DNA damage, and is effective at treating tumors that have already developed resistance to other cytotoxic treatments such as radiotherapy, hormone therapy, or chemotherapy (10, 18–22).

Despite these advantages, one major drawback of PDT is the limited penetration depth of light. PDT agents generate reactive oxygen species after the interaction with light, and the wavelengths of light for most of the clinically approved photosensitizers are in the ultraviolet (UV)/visible range (23, 24). This limits the use of conventional PDT methods to skin (surface) tumors only, and it is not effective for deep tumors (5, 17, 25–28). Recently, possible solutions to treat deep tumors with PDT have been proposed, for example: (i) the use of agents activatable by near-infrared (NIR) light with relative longer wavelengths (29–38), (ii) the use of upconversion nanoparticles that absorb NIR light and emit visible light to activate conventional photosensitizers (39–42), (iii) the use of fiber optics that transmit light deep into tissue (37), and (iv) the use of ionizing X-rays for photosensitizer activation (27). However, NIR light with enough energy to activate photosensitizers can penetrate only 5 mm into tissue (43). Similarly, upconversion nanoparticles are also limited by the penetration depth of NIR. The use of fiber optics is invasive, inconvenient, and cannot effectively and homogeneously activate the photosensitizers (44, 45). Furthermore, the treatment

of metastatic sites or lymph nodes is difficult as these sites are located in regions where light delivery is challenging. In contrast, the use of X-rays to activate photosensitizers may overcome the challenges of light penetration as X-rays, already used in medical imaging and therapy, can easily penetrate as deeply as necessary into patients.

Copper-cysteamine (Cu-Cy) nanoparticles are promising photosensitizing agents that can be effectively activated by X-rays to produce singlet oxygen for efficient deep cancer treatment (46, 47). Fig. 1 shows a schematic of the use of Cu-Cy nanoparticles for X-ray-activated photodynamic therapy. In this paper, we expand on a previous pilot study to measure the survival of a large cohort of mice after X-ray activation of Cu-Cy nanoparticles in comparison with controls. In this study, Cu-Cy nanoparticles are conjugated to a pH-low insertion peptide (pHLIP) (48–51) to facilitate active targeting of these nanoparticles to low pH tumors in the future.

## Results

**pHLIP-Cu-Cy Nanoparticle Characterization and Assessment.** As energy transfer is needed to produce singlet oxygen using X-rays, nanoparticle luminescence will occur, with stronger luminescence potentially resulting in more effective singlet oxygen production, and more efficient PDT. Cu-Cy nanoparticles exhibit strong luminescence, while many copper complexes have no luminescence due to efficient internal conversion (52, 53). Fig. 2A shows the Cu-Cy nanoparticles suspended in an aqueous solution, and its fluorescence under UV excitation is shown in Fig. 2B. The TEM image demonstrated that the average size of Cu-Cy

## Significance

Copper-cysteamine nanoparticles can be activated directly by X-rays to produce singlet oxygen. The use of copper-cysteamine nanoparticles (conjugated with pH-low insertion peptide) can enhance the effects of X-ray-induced photodynamic therapy, to lead to improved tumor treatment in mice. The results of this study demonstrate the potential of copper-cysteamine nanoparticles with deeply penetrating X-rays in the treatment of mammalian cancer to overcome current limitations of low penetration, light-induced photodynamic therapy treatment that can only treat superficial cancers.

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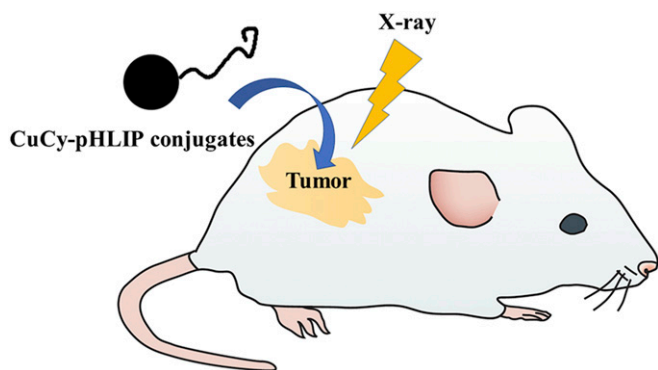
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**Fig. 1.** A schematic illustration of X-ray-induced PDT with pHLIP-conjugated Cu-Cy nanoparticles in mice.

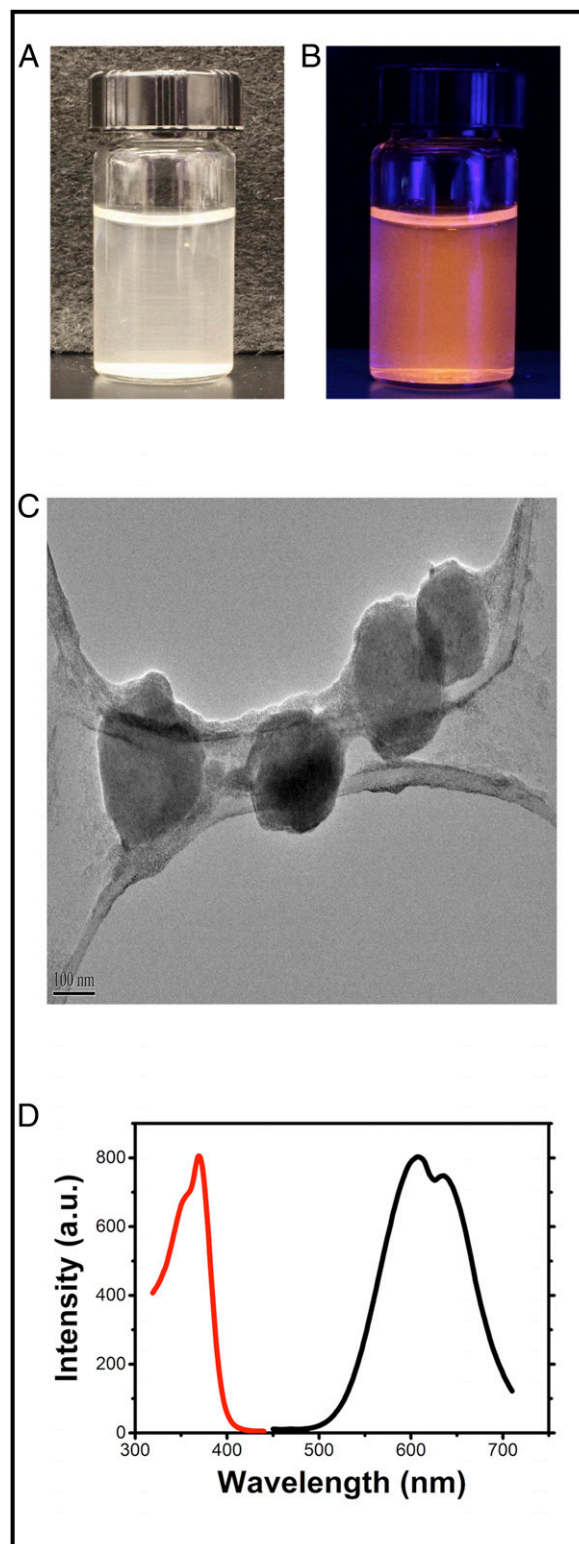
nanoparticles was ~200 nm. In Fig. 2C, the Cu-Cy nanoparticles appear to have a flake structure surrounding them, which are likely the seeds of Cu-Cy nanoparticles because Cu-Cy has a layered crystal structure, and it may form some flake-like crystal seeds. Upon excitation at 365 nm, these nanoparticles demonstrated emission in the red, peaking at 607 nm with a shoulder at 633 nm, indicating two luminescence-emitting centers (Fig. 2D). The luminescence of Cu-Cy corresponds to the Cu MC transition ( $d^94s^1-d^{10}$ ), which can be strongly affected by Cu-Cu interactions (54). These two emission peaks from Cu-Cy are caused by two types of copper ions, Cu (1) and Cu (2), existing in the Cu-Cy nanoparticles, which are different from each other by different coordination (55).

The conjugation of poly(ethylene glycol) methyl ether thiol-coated Cu-Cy nanoparticles with pHLIP were characterized by optical absorption and luminescence spectra (56, 57). Cu-Cy nanoparticles have only red emission with doublet peaks at 606 and 636 nm. However, the PHLIP-Cu-Cy nanoparticle conjugates have two emissions in the blue and the red (SI Appendix, Fig. S1). The red is from Cu-Cy nanoparticles, and the blue emission at 448 nm is due to the peptides. Cu-Cy nanoparticles have a strong absorption at 360 nm (SI Appendix, Fig. S2). When the conjugates were excited at 360 nm, both emissions from Cu-Cy nanoparticles and the peptides are observed. When the excitation spectrum was recorded by monitoring the blue emission at 448 nm, both the excitation peaks at 280 and 360 nm were observed. This indicated there is energy transfer between Cu-Cy nanoparticles and the peptides and that the conjugation was successful.

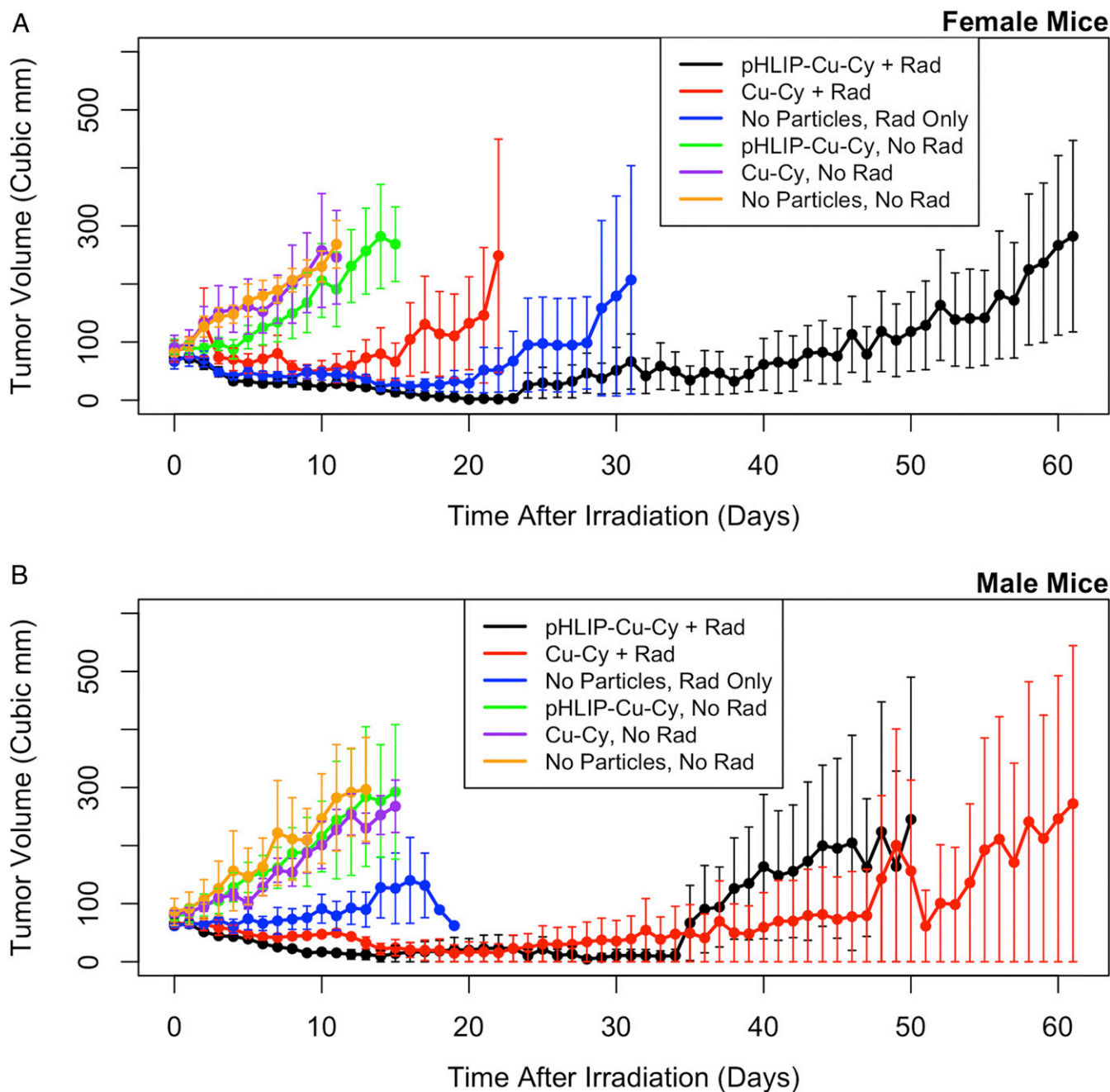
#### Effect of Cu-Cy Nanoparticles on Tumor Size after Radiation Therapy.

Fig. 3 and SI Appendix, Fig. S3 shows the average tumor volume as a function of time after irradiation. pHLIP-conjugated Cu-Cy nanoparticles produced a reduction in tumor size in both sexes, compared with mice given Cu-Cy nanoparticles (not conjugated to pHLIP) and radiation, mice given radiation only (no nanoparticles), and all nonirradiated mice. Longitudinal data analysis in the joint model (SI Appendix, Table S1) shows that radiation ( $P = 0.0001$ ) and pHLIP-conjugated Cu-Cy nanoparticles ( $P = 0.0386$ ) had significant negative effects on tumor size, indicating that both radiation therapy and pHLIP-conjugated Cu-Cy nanoparticle treatment tend to reduce the tumor size (cube root of volume) during the course of the study. In addition, the baseline tumor size ( $P < 0.0001$ ) had a significant positive effect, meaning that mice with larger tumor sizes at the initial time point were more likely to have a larger tumor size at the later time points. The quadratic term of time effects ( $P < 0.0001$ ) found to be significant is known to have an explicit effect on tumor size as a function of time. The effect of sex was found to be statistically

insignificant ( $P = 0.5091$ ). To control the familywise error rate (FWER = 5%) (58), a Bonferroni correction was applied (SI Appendix, Table S1), and the difference between radiation therapy with pHLIP-conjugated Cu-Cy nanoparticles and



**Fig. 2.** A photo of the Cu-Cy nanoparticles in aqueous solution (A), under a UV lamp (B), and under TEM (C). (D) The excitation (emission at 645 nm, Left) and emission (excitation at 365 nm, Right) of Cu-Cy nanoparticles.



**Fig. 3.** Tumor size as a function of time for female (A) and male (B) mice. Treatment with pHLIP-conjugated Cu-Cy nanoparticles and radiation shows a reduction in tumor size, compared with treatments of plain Cu-Cy nanoparticles and radiation and treatments of radiation only (no nanoparticles). Mean and SEM is plotted. Curves in this figure were cut off when average tumor size reached 300 mm<sup>3</sup> for the first time, or when only one mouse was left alive. For the last two data points of the radiation-only mice (blue), the error bars are too small to be visible. The full dataset used in the analysis is plotted in *SI Appendix, Fig. S3*.

radiation therapy with no nanoparticles was negatively significant ( $P < 0.0001$ ), indicating that pHLIP-conjugated Cu-Cy nanoparticles can significantly reduce the tumor size under X-ray activation compared with X-rays alone.

In contrast to the results for pHLIP-conjugated Cu-Cy nanoparticles, the difference between the treatments (radiation therapy and plain Cu-Cy nanoparticles versus radiation therapy with no nanoparticles) was not statistically significant ( $P = 0.5082$ ). This shows that tumor size did not significantly decrease in this experiment when irradiated with plain Cu-Cy nanoparticles, suggesting a benefit from the pHLIP conjugation. Further supporting this conclusion, the difference between

the treatments (radiation therapy and pHLIP-conjugated Cu-Cy nanoparticles versus radiation therapy and plain Cu-Cy nanoparticles) was statistically significant ( $P < 0.0001$ ), with the pHLIP-conjugated nanoparticles resulting in smaller tumor sizes. The results of survival data analysis in the joint model are given in *SI Appendix, Table S1*, where no covariates were found to be significant.

#### Discussion

In this paper, we demonstrated that Cu-Cy nanoparticles conjugated to pHLIP can reduce tumor size when combined with radiation therapy in mice. Cu-Cy nanoparticles can be used in



the treatment of both shallow and deep tumors because it can be activated by X-rays as well as light (46, 47). Cu-Cy nanoparticles interact with X-rays and produce both fluorescence (Fig. 4A) and singlet oxygen (55). Singlet oxygen is a reactive oxygen species, which causes damage to cells. Fig. 4B shows the energy level structure of Cu-Cy nanoparticles; in particular, the intersystem crossing from  $S_1$  to  $S_0$  results in an energy transfer and singlet oxygen generation. The mechanism for X-ray interactions with Cu-Cy nanoparticles to produce singlet oxygen is similar to the process for light activation to produce singlet oxygen; the only difference is that the excitation with X-rays is to higher excited levels, while the relaxation to lower energy levels and the energy transfer from the triplet state to excite dioxygen to produce oxygen are the same as illustrated (Fig. 4B). Cu-Cy nanoparticles have been used in therapy for SW620 colorectal cancer (47) and MCF-7 cells both in vitro and in vivo (46).

pHLIP-conjugated Cu-Cy nanoparticles showed the enhanced radiation effect with improved tumor size reduction in both male and female mice. It is possible that pHLIP contributed to the enhancement effect by binding Cu-Cy directly to a cell. Singlet

oxygen is known to have a short lifetime ( $\sim 4 \mu\text{s}$ ; refs. 59 and 60); thus, it may be of additional use to connect Cu-Cy directly to a cell. The effect of initial volume in the longitudinal analysis makes logical sense, because a tumor starting at a larger volume will have larger volumes overall than if it had started at a smaller volume. However, it is still a reminder of the importance of having mice irradiated with initial tumor volumes grouped as closely as possible.

Two particularly important variables that were not tested in this work are radiation energy and radiation dose. Few, if any, photoluminescent particles have been shown to work at energies as high as 90 kVp, which was the radiation energy spectrum applied in this work. However, most clinically relevant energies are higher. In the future, the effectiveness of higher energy photons and delivered dose will be evaluated. In addition, although this paper is about the application of PDT to deeper cancers, many skin cancers are treated with kiloelectron volt-level radiation (61), and Cu-Cy nanoparticles may be able to enhance the effectiveness of this process.

Overall, this paper demonstrates the strong potential of pHLIP-conjugated Cu-Cy nanoparticles, combined with X-rays, as a photosensitizer for PDT to successfully treat mammalian cancer.

## Materials and Methods

**Cu-Cy Nanoparticle Synthesis and Characterization.** Cu-Cy nanoparticles were synthesized at The University of Texas at Arlington (56). For size characterization, Cu-Cy nanoparticles dispersed in water were placed on holey carbon-covered copper grids for HRTEM observations. The HRTEM images of the particles were obtained with a Hitachi 9500 electron microscope using an accelerating voltage of 300 kV (55).

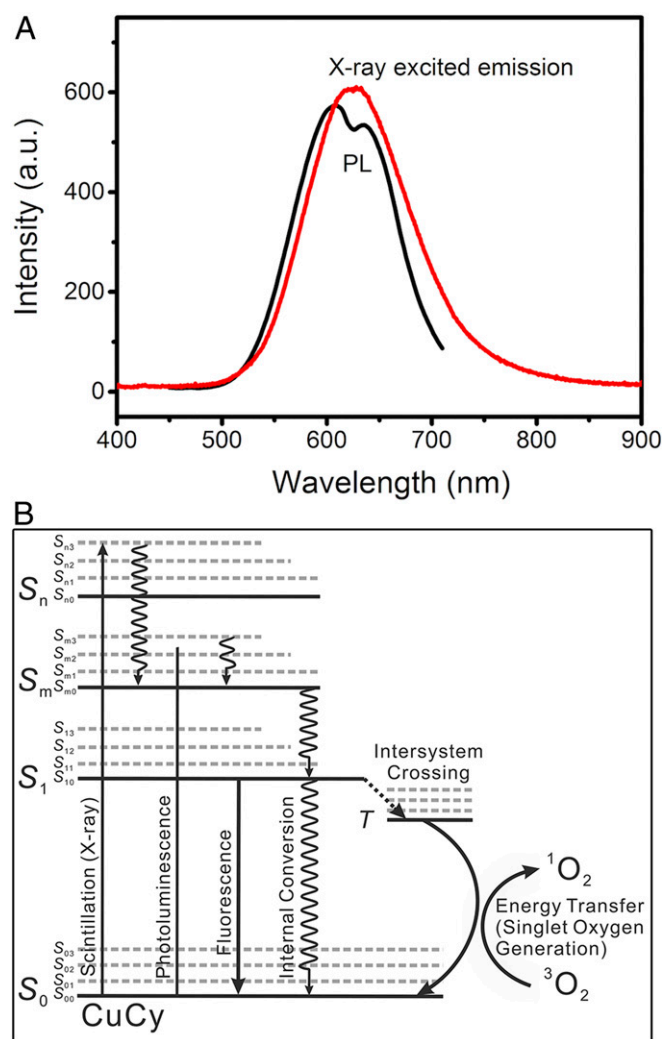
**Preparation of pHLIP Conjugated Cu-Cy Nanoparticles.** Two milligrams of Var3 pHLIP (Ala-28-Gly), from CS Bio Company, was added to 5 mL of deionized water followed by the addition of 3.19 mg of 1 Ethyl-3-(3-dimethylaminopropyl) carbodiimide under mild stirring for 10 min at room temperature. After adjusting the pH to 7.5 using NaOH, 5 mL of 1 mM Cu-Cy nanoparticles (in a water solution) was added under constant stirring overnight at room temperature in a dark environment. The pHLIP-Cu-Cy nanoparticle conjugates were centrifuged at 4,400 rpm for 25 min and washed with deionized water three to four times to remove the precipitates and then were purified by dialysis to remove unreacted species.

**Cell Culture.** JC Breast murine cancer cells of BALB/cRos strain (CRL-2116) were purchased from American Type Culture Collection and were grown in Roswell Park Memorial Institute (RPMI) medium with L-glutamine and sodium bicarbonate, 10% FBS (Sigma-Aldrich), and 0.1% Ciprofloxacin. The cells were maintained in a humidified atmosphere at 5% carbon dioxide at 37 °C in an incubator. Breast cancer is one example of a cancer type that is reachable at the X-ray energies used in this experiment.

**Animal Models and Cell Injection.** All animal work followed the guidelines of the University of Rhode Island Institutional Animal Care and Use Committee protocol AN1516-003. Male and female BALB/c mice, 3–4 wk in age, were ordered from Envigo. Male and female mice were used to take the important biological variable of sex into account. For tumor cell inoculation, 1.5 million cells were suspended in 100  $\mu\text{L}$  of RPMI and injected subcutaneously on the right flank of the mice using 1 mL of 27 G<sup>1/2</sup> latex-free BD syringes.

**Radiation Therapy on Mice.** Mice were divided into six treatment groups: (i) pHLIP-Cu-Cy nanoparticles + radiation; (ii) Cu-Cy nanoparticles + radiation; (iii) phosphate-buffered saline (PBS) + radiation; (iv) pHLIP-Cu-Cy nanoparticles; (v) Cu-Cy nanoparticles; and (vi) PBS (control). In total, 51 mice (24 males and 27 females) were used for the experiment.

Treatment was undertaken when the tumor size (length) reached  $\sim 4\text{--}8 \text{ mm}$ . Mice were anesthetized using isoflurane gas. For groups of mice given nanoparticles, the nanoparticles were injected intratumorally in 20  $\mu\text{L}$  of PBS at a nanoparticle concentration of 0.8  $\mu\text{g}/\mu\text{L}$ . For the groups given radiation therapy, the mice were irradiated 30 min after injection of particles with an irradiation dose of 5 Gy. Mice were shielded using lead; the only area irradiated was the vicinity of the tumor—the irradiation area was an approximate semicircle 1 inch in diameter. No external X-ray filter was used, and the



**Fig. 4.** (A) Photoluminescence (PL, excitation at 365 nm) and X-ray-excited luminescence spectra from Cu-Cy nanoparticles. (B) Schematic illustration of the scintillation processes in Cu-Cy nanoparticles upon X-ray irradiation. After X-ray irradiation, it may produce photoluminescence, fluorescence, internal conversion, and an intersystem crossing that transfers energy to excite oxygen to produce singlet oxygen.

source to surface distance was set to 30.5 cm with a field size of 18.3 by 20.1 cm. The current and voltage settings of the X-ray machine (Faxitron MultiRad 350) were 90 kVp and 30 mA, respectively. The nonirradiated mice were also placed in the X-ray chamber, with the same settings except no radiation was given. The tumor size was measured daily using digital Vernier calipers (VCD001, from United Scientific Supply) to get the tumor volume. The tumor volume was calculated using the formula tumor volume =  $1/2 \text{ length} \times \text{width}^2$  (62). Mice were euthanized if they reached or approached the endpoint tumor length of 20 mm, the tumors were necrosed, or if the mice showed signs of distress. Nine mice were euthanized before these endpoints were reached (between days 57 and 93 after irradiation); one mouse was found dead during the experiment.

For tumor targeting with pHILIP, pHILIP-conjugated Cu-Cy nanoparticles should be injected intravenously (i.v.) instead of intratumorally. As a first step, intratumoral injections were used to more accurately compare these results with previous work (46). After this study, i.v. injections will be evaluated in future work.

**Statistics and Analysis of Data.** In total, 51 mice were used: 24 males and 27 females. Each of the radiation therapy groups had three males and four females, whereas the nonradiation therapy groups had five males and five females. Given that the data are longitudinal with follow-up truncated by death (63), a joint model of longitudinal (cube root of daily tumor volume) and survival response (time to death) was applied to assess the longitudinal trajectory effect of each treatment and its impact on survival simultaneously while controlling for contributing baseline and demographic factors. The effects of baseline (tumor size at irradiation) and demographic (sex and estimated age at irradiation) factors were controlled by including those

factors as independent variables/covariates in the joint model. The cube root of volume was used as a measure of tumor size (64, 65). This approach is similar to the concept of the geometric mean of the tumor size (cubic root of length times width times height; ref. 66), although the tumor height was not measured here.

A mixed-effects regression model was assumed for the longitudinal data, and a Cox proportional hazards model was assumed for the survival data. To induce the correlation between longitudinal and survival response, we further assumed the mixed-effects regression model and the hazard function of Cox model shared the same quadratic time trajectory function (67). The covariates considered in both longitudinal and survival models include sex, radiation, treatment effects (no nanoparticles, pHILIP-Cu-Cy nanoparticles, and Cu-Cy nanoparticles), interaction effects between radiation and treatment, age at irradiation, and tumor size at irradiation (cube root of volume). Multiple comparisons between different treatments were also conducted. *P* values from multiple comparisons were corrected by the FWER approach. All analyses were run in SAS version 9.4 using JMFIT SAS macro.

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